Using Oxygen Isotope Analysis of Modified Cellulose to Investigate Salt Water Intrusion and Paleotemperatures

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INTRODUCTION- Organic Oxygen Isotopes

- ¹⁸O vs. ¹⁶O
 - .2% of oxygen atoms vs. 99.76% of oxygen atoms
- Factors affecting the ¹⁸O content of organic matter
 - Variation in the source water $\delta^{18}O$
 - Leaf water evaporative δ¹⁸O enrichment
 - Water mixing within the plant (stem/leaves)
 - Isotope exchange between water and organic oxygen

INTRODUCTION- Source Water δ^{18} O

- $\delta^{18}O_{rainfall} = 0.52T 0.006T^2 + 2.42P 1.43P^2 (0.046Elv^{1/2})$ -13.0
 - T = temperature, P = precipitation, E = elevation
 - Higher temperatures = higher δ^{18} O values
 - More precipitation = lower δ^{18} O values
 - Higher elevations = lower δ^{18} O values
 - Barbour et. al (2001), based on data from IAEA (1992)
- Because of the relationship to temperature, δ18O values of cellulose have been proposed as "isotopic thermometers"
 - Libby et. al 1976
- Groundwater vs. Precipitation
 - Different δ^{18} O signatures, Clark and Fritz, 1997.

INTRODUCTION- Leaf water δ^{18} O

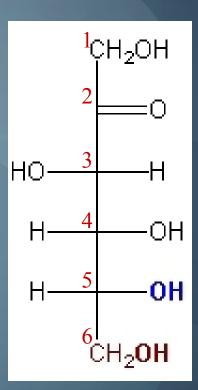
- Because ¹⁶O atoms evaporate more readily, leaf water gets enriched according to the Craig-Gordon equation (1965):
 - $\delta^{18}O_E = \delta^{18}O_s + \varepsilon^* + \varepsilon_k + e_a/e_i \left(\delta^{18}O_A \delta^{18}O_S \varepsilon_k\right)$
 - $\delta^{18}O_s$ = isotopic composition of the stem water
 - ϵ^* and ϵ_k =
 - \mathbf{e}_{a} = ambient vapor pressure/ \mathbf{e}_{i} = intercellular vapor pressure
 - e_a/e_i = relative humidity, leaf is usually saturated
 - $\delta^{18}O_A$ = isotopic composition of atmospheric water vapor
- Transpiration reduces leaf water evaporative enrichment by drawing stem water into the leaf.
 - Péclet Effect equation: $\rho = (TL)/(CD)$
 - T is transpiration rate, L is the mixing length of the leaf
 - C and D are constants.
- Combined equation predicts bulk leaf water enrichment.
 - $\Delta^{18}O_{l} = [(1-\alpha)\delta^{18}O_{s}] + [\alpha\delta^{18}O_{E}]$
 - $\alpha = (1 (1/e^{\rho}))/\rho$

INTRODUCTION- Mix of Source and Leaf Water

- Phloem transports sugar and water from the leaves to growing cells.
 - Slightly less enriched than leaf water, because it equilibrates with source water in the xylem
 - Bret-Harte and Silk, 1994
- The two water sources mix; how they mix effects the δ^{18} O of the cellulose.

INTRODUCTION- Biochemical fractionation

- To make cellulose, sucrose breaks into glucose and fructose.
 - Sucrose has δ¹⁸O of leaf water (Barbour, 1999)
- Glucose and fructose exchange in equilibrium.
- The oxygen atom on the carbonyl group (C=O) exchanges with the oxygen in the water.



Fructose

INTRODUCTION- Biochemical noise

- Some fructose molecules are used by the plant to make other, non-cellulose compounds. Lighter fructose molecules with ¹⁶O on the second carbon are used first, changing the isotopic ratio.
 - Example: manitol
- Phenolglucosazone removes the oxygen from the first and second carbons, eliminating the oxygen isotope changes caused by fructose use.
 - Sternberg et. al., 2006
- Even without considering that particular biochemical factor, δ^{18} O ratios are affected by many different variables and can be difficult to interpret correctly.

INTRODUTION- Conocarpus erectus

- aka Buttonwood
- Survive at up to 1,000 ppm
 NaCl
- Salinity vs. $\delta^{18}O$
 - More salt more ¹⁸O
 - Sternberg et. al, 1991
- Hypothesis- Buttonwood samples collected from freshwater areas will have lower cellulose and PG δ^{18} O values than those collected from saltwater areas.
- Application- Buttonwoods could provide a record of salt-water intrusion



INTRODUCTION- Sphagnum

- aka peat moss
- Sphagnum genus of bryophyte
 - Leafy stem topped with capitulum, stem decays while plant keeps growing
- Slow decay rate preserves organic matter
 - acidic conditions, phenolic compounds, dry/cold conditions
- Hypothesis- The δ^{18} O values of derived phenolglucosazone will be more strongly correlated with estimated paleotemperatures than the δ^{18} O of peat cellulose.
- Application- Peat cores are often used to establish paleoclimate conditions, including temperatures.

METHODS

- Sample Collection
 - Buttonwood samples collected from University of Miami, SW 62nd Avenue, Matheson Hammock, Sugar Loaf Key
 - Peat cellulose samples sent by Elise G. Pendall, U of Wyoming
- Cellulose Extraction
 - to isolate cellulose from bulk dry plant matter
 - Leavitt and Danzer, 1993
- NaOH wash
 - to isolate α-cellulose, eliminate hemi-celluloses
- Hydrolysis
 - to break up cellulose chains into glucose monomers
 - Fengel and Wegener, 1979
- Derivation
 - to form phenolglucosazone from glucose
 - Oikawa, 1998
- Isotope Ratio Mass Spectrometer (IRMS)

RESULTS

Cellulose

y = -.0076x + 30.767

 $R^2 = .0001$

(no relationship)

Phenolglucosazone

y = .2151X + 22.168

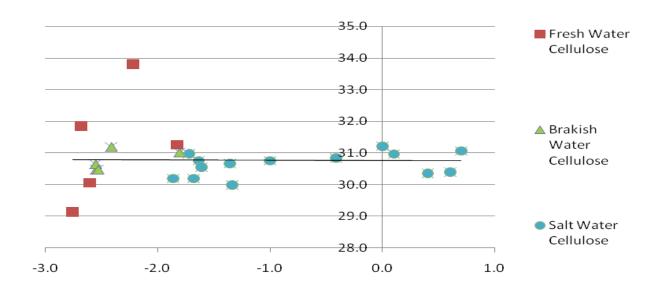
 $R^2 = .0306$

(no relationship)

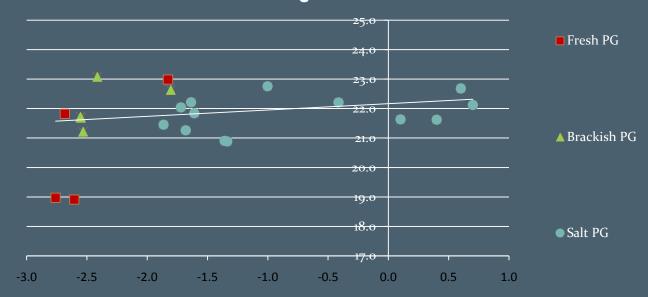
Neither cellulose nor phenolglucosazone does a very good job of predicting stem water $\delta^{18}O$

Neither result is statistically significant.

The Effect of Stem Water δ18O on the δΟ18 of Collected Cellulose



The Effect of Stem Water δ018 on the δ180 of Collected Phenolglucosazone



DISCUSSION

Conocarpus erectus

- δ^{18} O of cellulose unrelated to δ^{18} O of stem water, same for phenolglucosazone
- δ^{18} O of cellulose almost constant
 - Difficult to use for study of salt-water intrusion

Sphagnum/Peat

- Peat Samples did not produce phenolglucosazone.
- Fresh Sphagnum moss did produce phenolglucosazone.
 - Low yield
- Possible presence of molecules in peat strongly bonded to cellulose which may interfere with phenolglucosazation.
 - lignin?

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